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## Effects of valproic acid on rat C6 glioma cells proliferation and animals survival in the experiment

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**Background.** Treatment of glioblastoma is an extremely important problem of neuro-oncology. Despite the presence of modern medical and technological developments, there is no significant progress in solving it today. Valproic acid (VPA) is an antiepileptic drug with proven efficacy in epilepsy, and potential oncostatic effects of VPA in the treatment of brain tumors are considered. This study examined the effect of VPA on the growth of rat C6 glioma *in vivo*

**Methods.** After experimental glioma modeling in rats, valproic acid injections were performed intraperitoneally. Survival of rats was studied according to the Kaplan-Meier curve and tumors were examined histologically. Conclusions about proliferative activity were made based on the determination of the index of Ki67-positive cells

**Results.** Valproic acid significantly increased the median survival rate of rats with gliomas from 11 to 13 days ( $p=0,05$ ) and significantly decreased the proliferative activity of glioma cells ( $3.53\pm 0.96$ ,  $Me=3.08$  vs  $2.17\pm 0.38$ ,  $Me=2.11$ ,  $p=0,05$ )

**Conclusions.** These findings indicate that valproic acid inhibits the growth of glioma cells *in vivo*, which can be considered as a promising tool in the complex therapy of gliomas in clinical practice and is a perspective for further research

**Key words:** rat C6 glioma; survival; valproic acid (VPA); proliferation

## Introduction

Glioblastoma is a tumor that originates from glial cells of the central nervous system, characterized by aggressive malignant growth, a high rate of recurrence and high mortality rate of patients [1].

Increasing the survival rates and improving the quality of life for patients with glioblastoma is an extremely urgent problem of neurosurgery, since the average life expectancy of such patients is 14.6 months, and the five-year survival rate in developed countries does not exceed 6.3%. Therefore, scientific research and optimization of both surgical techniques and adjuvant treatment methods remain highly relevant: over the last 25 years the life expectancy of patients has increased with glioblastoma by only 3.3 months [2, 3]. Survival rates are one of the most significant **criterion** in oncology. They are used for the most adequate evaluation of the effectiveness of various treatment methods in clinical studies, evaluation of the medical care organization effectiveness for the population in oncoepidemiological population studies [4] in experimental medical and biomedical research [5]

Kaplan-Meier (KM) **survival analysis** is a non-parametric estimate of the survival function that

is commonly used to describe the survival of a study population and to compare two study populations [6]. This indicator is a reliable and convenient tool for comparing the life expectancy of patients (or experimental animals) when investigating new treatment methods compared to a control group (standard treatment). Thus, by comparing two or more survival curves, you can get a visual conclusion about the effectiveness of experimental treatment. The essence of such a comparison is to test null hypothesis - the assumption that survival in the two groups does not differ and the studied factor (treatment method) does not affect the studied value (survival rates). If there is a significant difference in the survival of the two studied groups, null hypothesis is rejected, which will indicate the effectiveness of a new or experimental method of treatment [7].

Animal models of tumors are valuable for studying the biological features of tumors and identifying agents with a potential oncostatic effect. The C6 glioma cell line was isolated from Wistar-Furthi rats, and is of astrocytic origin is both a well-characterized and most widely used glioma model, given its high malignancy and aggressiveness [5]. Rat glioma cells express a similar profile of human glioma markers (PDGF $\beta$ , IGF-1, EGFR,

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Erb3/Her3) [8]. C6 glioma is a model of glioma cell proliferation and migration and is used to evaluate the potential antitumor growth of new drugs [5, 8].

The use of antiepileptic drugs in clinical practice (in the complex therapy of gliomas) is natural, since the frequency of symptomatic epilepsy can reach 100%, and in particular in glioblastoma - up to 60% [9]. Valproic acid, an antiepileptic drug from the valproate group, is primarily indicated for various forms of epilepsy, migraine management, bipolar disorders, etc. It also exhibits antipsychotic effects [10] and acts as a radiosensitizer in glioma treatment [11]. The realization of the clinical effects of VPA is carried out through its various mechanisms: blockade of sodium and calcium channels of cell membranes, as well as an increase in the concentration of GABA in brain tissues [12]. The oncostatic effect of VPA has been described in many types of tumors: gastric and breast cancers, etc., research data is still ongoing [13, 14], and is relatively new. Regarding glioblastoma, this effect of VPA has been known since the early 2000s, when the effect of histone deacetylase inhibition was discovered, which led to a decrease in mitotic activity in tumor cells and, as a result, to a decrease in its progression [6, 15, 16]. Data from experimental studies *in vivo* and *in vitro* on inhibition of glioblastoma growth under the influence of VPA show encouraging results [17, 18, 19], but the results of clinical use are quite contradictory and ambiguous: it is reported about a positive experience of using VPA in glioblastomas and anaplastic astrocytomas [11, 10], especially in children and young people, while it is completely ineffective for neuro-oncology patients [20].

Thus, the search for new treatment methods and the potential effect of VPA on glioma remains relevant. In this study we focused on investigating the effects of valproic acid on survival of laboratory animals using the well-characterized rat brain C6 glioma cell line, aiming to establish the potential impact of valproic acid on the proliferative activity of glioma cells *in vivo*.

**Aim of the study:** To determine the effect of valproic acid on the proliferative activity of C6 rat glioma cells based on the analysis of Ki67 indicators and the survival of animals with glioma. To determine the prospects and feasibility of further studies of the oncostatic effect of VPA in the treatment of glial tumors, based on the results of this experiment and literature data.

## Materials and methods

### C6 glioma model

White outbred male rats weighing  $100 \pm 25$  g, with an average age of 2 months, were selected for the study. The animals were kept in the vivarium under the conditions of a natural circadian light cycle. They were fed a combined balanced diet *ad libitum* and with free access to water. The control group was 10 animals and the experimental group was 14 animals. To transplant glioma cells from a sick animal, a tumor was removed from which a cell suspension was prepared. After counting the cells in the Horyaev chamber, all animals were implanted with C6 rat glioma cells ( $2 \times 10^6$  cells in

0.1 ml) in the right temporal area to a depth of 4 mm, as described [21].

This study was conducted in strict accordance with the recommendations of the National Institutes of Health's Guide for the Care and Use of Laboratory Animals. The research protocol with animal experimentation was approved by the Scientific Ethics Committee (Protocol Number: 17/2022). All surgery was performed under general anesthesia and every effort was made to minimize suffering. The study was conducted on two groups of animals with glioma modeling: 1 - control; 2 - experimental, which was given VPA treatment.

### Surgical procedure

Manipulations with animals were carried out under general anesthesia - intraperitoneal injection of thiopental sodium 50-60 mg/kg of animal weight. The injection site was previously disinfected with a 5% iodine solution.

All surgical procedures were carried out using sterile instruments in operation room. Prior to surgery, animals were fixed in stereotactic device. Skull operative area, was decontaminated by betadine solution with a cotton swab. Coronal suture was exposed through midline scalp incision. Cranial burrhole was made 4 mm to the right of midline and 1 mm anterior to the coronal suture by a mini-drill. A 26-gauge stainless steel guide cannula was stereotactically implanted through this hole to a depth of 4 mm and the suspension of C6 rat glioma cells was injected ( $2 \times 10^6$  cells in 0.1 ml) in the right temporal area. The skin was closed using 7-0 (ETICON) atraumatic sutures.

The animals were observed daily and their lifespan was recorded. On the 5th day, one animal in the control group and on the 6th day one animal in the experimental group dropped out due to death, neither death was related to the tumor; in addition, on the 10th day, 6 animals were removed from both groups to study and compare the morphological features of the tumor. These animals were not taken into account in the calculation of survival rates due to identical elimination dynamics in the groups. Thus, the number of animals in the control and experimental groups was 10 and 14, respectively (**see Table 1**).

The preparation of valproic acid in the form of a solution of 100 mg/ml was administered intraperitoneally at a dose of 300 mg/kg of the animal's body once daily throughout the experiment. This dose was selected based on its safety profile and its lack of undesirable biological reactions [22]. Animals were removed from the experiment by injecting a lethal dose of sodium thiopental intraperitoneally.

**Table 1.** Number of animals

Group	Animals, N
Control	10
Experimental	14
Overall	24

*This article contains some figures that are displayed in color online but in black and white in the print edition*

### Histological examination

At 10<sup>th</sup> day of experiment, rat brains were fixed in a 10% formalin solution (pH 7.4, 4°C, 24 hours). Tumors were isolated from each brain, dehydrated in isopropanol and sealed in paraplast (Leica Surgipath Paraplast Regular). Sections with a thickness of 4 µm were obtained on a Thermo Microm HM 360 microtome. Sections after deparaffinization were stained with Sirius Red and Weigert's hematoxylin, hematoxylin and eosin to study tumor morphology. Proliferative activity was assessed by immunohistochemical method based on the analysis of immunopositive reactions to Ki67. (<https://www.abcam.com/en-us/products/primary-antibodies/ki67-antibody-sp6-ab16667#tab=datasheet>) A monoclonal antibody against Ki67 (Abcam, USA, Cat.number: ab 16667) was used. Primary antibodies were used at a dilution of 1:200. Reaction products were visualized using a diaminobenzidine-based detection system (EnVision FLEX; Dako, Glostrup, Denmark). Sections were incubated with antibodies at a temperature of +24°C (with primary and secondary antibodies for 20 and 10 min, respectively). Cell nuclei were stained with hematoxylin Gill I. The obtained micropreparations were examined in an Olympus BX51 microscope, magnification ×200, ×400. Proliferative activity was evaluated as the relative number of Ki67-positive cells in 10 fields of view for each tumor sample, as the number of Ki67 positive cells to the total number of cells in the region of interest.

### Statistical analysis

Data processing was carried out using the statistical software package Statistica 10.0 (StatSoft Inc., USA).

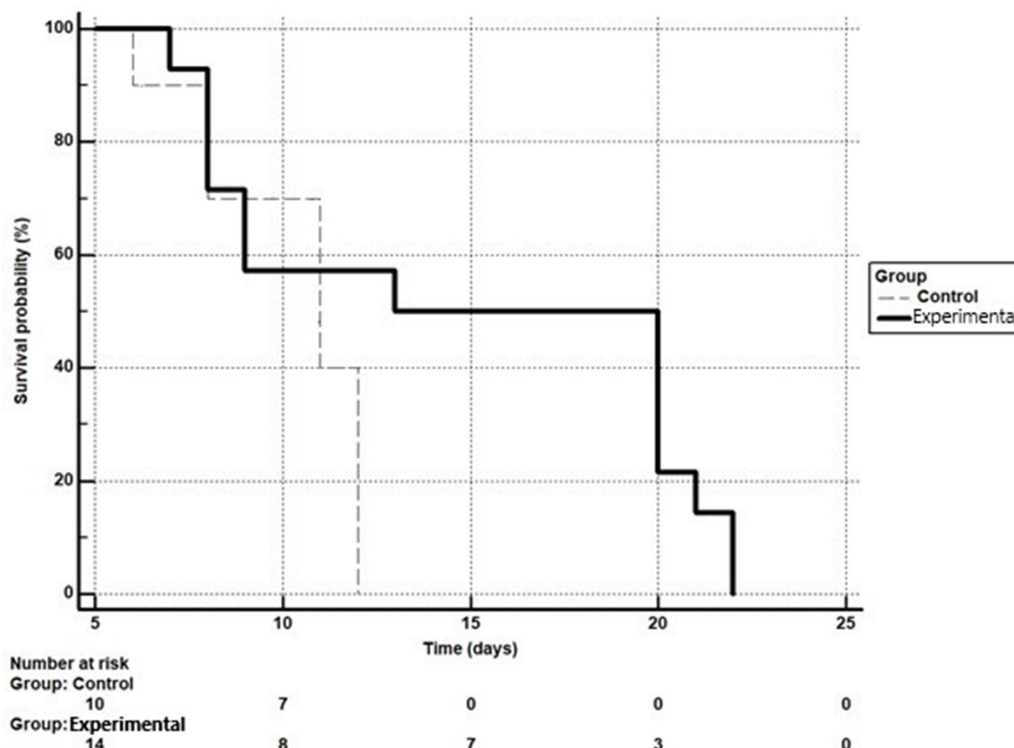
Survival rates were standardized, entered into the database and subjected to statistical processing using descriptive and non-parametric statistical methods. Survival analysis was performed using the Kaplan-Meier method. Pearson's  $\chi^2$  test was used to determine the difference between the survival of animals in the study and control groups. When comparing survival in different groups, the risk ratio with a 95% confidence interval (CI) was used. The Mann-Whitney test was used to analyze the difference in the values of proliferative activity between groups. The results were considered statistically significant provided that the level of statistical significance ( $p$ ) was  $<0.05$ . Spearman correlation coefficient was used to analyze the correlation between Ki-67 index and tumor mass. The results are given in the discussion.

## Results

### Survival analysis

In the control group, one animal died on the 6th day, two animals on the 8th day, three animals on the 11th day, and four animals on the 12th day. In the experimental group: on the 7th day – one animal, on the 8th day – three animals, on the 9th day – two animals, on the 13th day – one animal, on the 20th day – four animals, on the 21st day – one animal and on the 22nd day – two animals. Generalized data are shown in **Fig. 1**. The results of statistical processing of the given data with survival rates are shown in **Table 2**.

Thus, the median survival in the control group was 11 days, in the experimental group 13 days, which is demonstrated by the Kaplan-Meier curves in **Fig. 1**.



**Fig. 1.** Kaplan-Meier curve of survival of rats of the control and experimental groups. The difference between groups according to chi-square results is statistically significant ( $p=0.04$ )

**Table 2.** Survival rate of animals

Group	Mean survival (days)	SE	Confidence interval for the mean 95%	Median survival (days)	Confidence interval for the median 95%
Control	<b>10,300</b>	0,684	8,959 to 11,641	11,000	8,000 to 12,000
Experimental	<b>14,786</b>	1,691	11,472 to 18,100	13,000	8,000 to 20,000

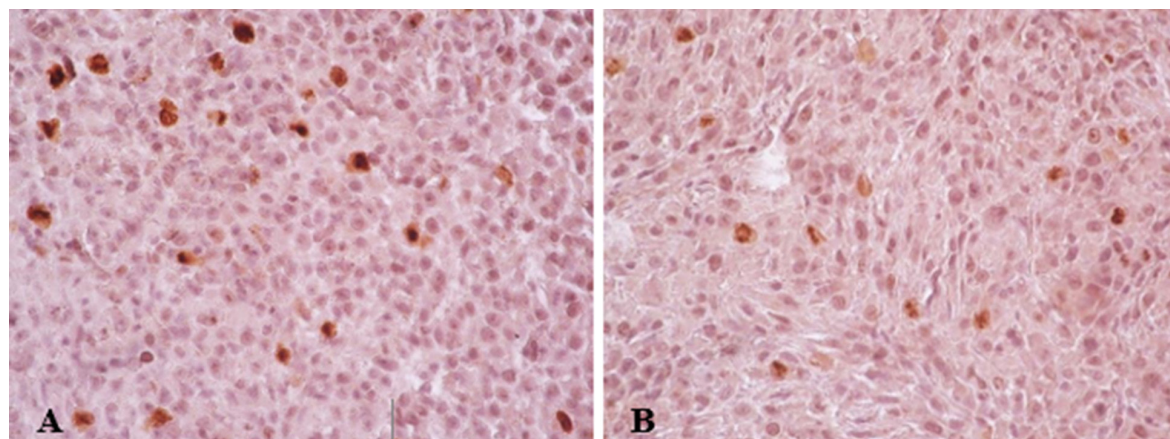
For the experimental group versus the control group: hazard ratio (HR)=0.51 (0.21-1.26), p=0.0448 – relative risk reduced by 49%. It is defined as (1-0.51)\*100. For the control group versus the experimental group: HR=1.96 (0.79-4.84), p=0.0448 – relative risk increased by 1.96 times.

The risk of death in the control group compared to the experimental group is 1.96 times higher (HR=1.96), but the risk of death in the experimental group compared to the control group is reduced by 49% (HR=0.51).

**Tumor morphology**

10 days after implantation of C6 glioma cells into the brain of rats, intracerebral tumors were detected. The tumors had a small volume and clear borders. This proved that the glioma model was successfully reproduced. The average weight of tumors was 254.1 ± 38.67 mg in the control group and 308.3 ± 55.8 in the experimental group (p>0.05). Glioma tissues in both the control and experimental groups were characterized by moderate nuclear atypia (nuclear pleomorphism) and not high mitotic activity. It was also found that a certain proportion of tumor cells in both groups had cytoplasmic vacuolation; this feature

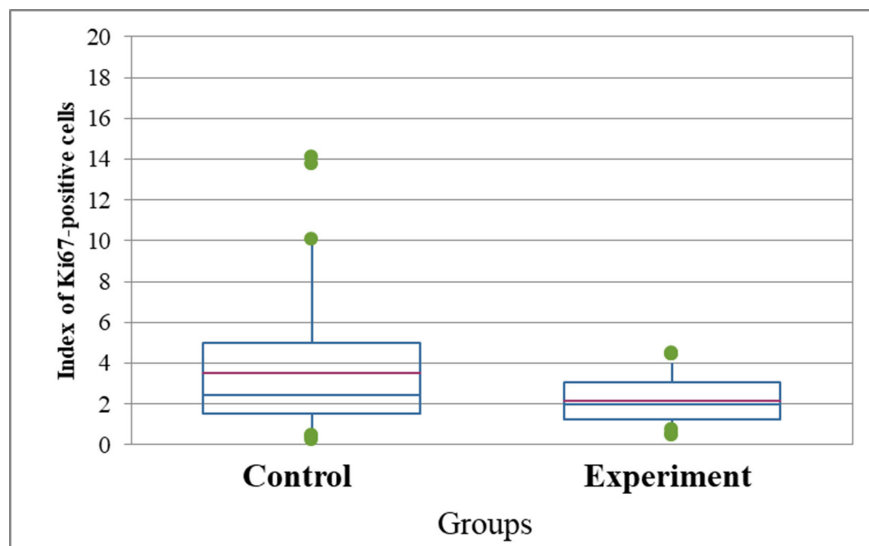
was slightly more prevalent in the experimental group. According to the results of staining with hematoxylin and eosin, Sirius Red, tumors grew intracerebral, glioma cells occasionally extended into the dura mater which is rich in collagen. The peripheral zone of the tumors had less vascularization than in the central part of the tumor. Areas of sparse necrosis were observed in the central part of the tumor contained only single Ki67-positive cells (**Fig. 2**). The Ki67 index in the control group was 3.5%, and in the experimental group it was 2.1% (**Fig. 3, Table 4**). According to the results of the assessment of the one-sided Mann-Whitney test, the difference was probable at p=0.01, and for two-sided p=0.02. Thus, the study results revealed a slight but statistically significant difference between the values of the proliferative activity index in the comparison groups. At the same time, a negative correlation was found between the mass of tumors and the average value of the proliferative index in the general sample of the experiment (r=-0.61, p=0.04), while for the control and experimental groups, the correlation analysis did not show a probable dependence (r =-0.71, p=0.11 vs r=-0.49, p=0.33)



**Fig. 2.** Micrographs of C6 glioma tumor tissues, Ki67-positive cells (brown). Control group (A) and the experimental group with valproic acid (B). Immunohistochemical detection of Ki67. Rev. 40, approx. 10

**Table 3.** Hazard ratios of survival with 95% confidence interval

Group	Hazard ratio of survival (HR) and 95%CI
Experimental vs Control	0,51 (0,21 to 1,26)
Control vs Experimental	1,96 (0,79 to 4,84)



**Fig. 3.** Index of Ki67-positive C6 cells in glioma tumors according to the results of Ki67 immunohistochemical detection (%). For the one-sided Mann-Whitney test  $p=0.01$ , for two-sided  $p=0.02$

**Table 4.** Index of Ki67-positive C6 cells in tumor tissues of control and experimental groups of rats on day 10 ( $n=24$ )

Group	Mean	SEM	Minimum	Q1	Median	Q3	Maximum
Control	3,53	0,96	1,29	1,45	3,08	5,51	6,48
Experimental	2,17	0,38	1,11	1,42	2,11	3,03	3,18

Notes. Mean - arithmetic mean value of the data in the group; Median - the average value from the main data array in the group; Q1 and Q3 are the interquartile range in the data set. All these indicators are used when conducting data analysis using the Mann-Whitney method, when the samples in both groups differ from the Gaussian distribution, making parametric methods inapplicable.

### Discussion

The rat C6 glioma model is widely used as an animal model in human glioblastoma research because it is similar in terms of growth and biological behavior to malignant brain gliomas [5, 23].

The results of *in vivo* research on the C6 glioma model in rats indicate a significant difference in both survival rates and **proliferative activity (Ki67) in glioma-bearing rats without treatment comparing to glioma-bearing rats with VPA treatment**, which correlates with literature data on this issue. In our opinion, a certain paradoxical situation has arisen regarding the effectiveness of VPA in the treatment of glial tumors: on the one hand, experimental data both *in vitro* and *in vivo* (and our own experience) convincingly testify to the oncostatic effect of valproic acid on glial tumors; on the other hand, there is no convincing clinical data on such effectiveness. This raises the question of prospects and possible directions of further research: possible ways to overcome the obvious difference between experimental data and clinical results can be implemented as follows:

1) increasing the dose of VPA to the maximum possible **level** for each specific patient [23] while exploring strategies to mitigate its toxic effect of VPA on the patient's body; for example, the use of various carbon nanomaterials (nanotubes, fullerenes, etc.) is promising for reducing toxicity [24];

2) combination with other medicinal products to potentiate their effect; for example, taking VPA in combination with temozolomide increased the median survival of patients with glioblastoma by 8 weeks, compared with the group where patients received temozolomide monotherapy [10, 22];

3) combination with other treatment methods, such as radiosurgery, radiochemotherapy and others [25, 11];

4) development of VPA-based drugs: separate experimental studies demonstrate that various VPA derivatives have different anticonvulsant, antitumor and histone deacetylase inhibition properties compared to VPA itself [26, 27, 28];

5) advancing drug delivery methods to target cells, for example, the use of nanoparticles containing an oncostatic agent (doxorubicin) has demonstrated a 40% cure rate in laboratory animals [29].

Another unsolved question that arises from the results of this work is the absence of a positive correlation between tumor mass and Ki67 as an indicator of proliferative activity of tumor C6 cells. In other studies in the C6 rat model, such a strong relationship was demonstrated between Ki67 values and mean tumor size measured by spectral CT [30] and studying the mechanisms of tumor cell death.

This issue requires further analysis and research by other methods, in particular, the detection of cellular markers of cell death.

Therefore, further studies of the VPA action in neuro-oncology are essential. Evaluating the impact of VPA on malignant cells suggests several possible mechanisms for its oncostatic action: a direct effect on the mitotic cycle by inhibiting histone deacetylase and an indirect one - through the inhibition of angiogenesis, activation of aging mechanisms and death of tumor cells. Cell death is typically caused by apoptosis or necrosis. However, an alternative scenario may occur - autophagy. It is already known that VPA is able to initiate a moderate apoptotic response by preferential activation of the mitochondrial pathway in cancer cells [31] and induce cell death through the autophagy pathway [32]. Microscopic differences in autophagy in cells include the detection of large vacuoles in the cytoplasm and specifically in the presence of VPA. In our study, vacuolization was not quantified as the primary focus was mainly aimed at an integral assessment of the oncostatic effect - that is, overall survival. However, the weak signs of vacuoles in the cytoplasm of cells in the VPA group suggests that autophagic cell death induced by VPA may be involved in the tumor growth inhibition. VPA was also found to disrupt blood vessel formation by reducing eNOS expression [33]. In general, given the diversity of VPA effects on biological systems, at the level of macromolecules, cells, tumor tissue, and the organism as a whole, the molecular mechanisms of the oncostatic action of VPA require further research.

### Conclusions

1. A significant increase in survival rates and median survival was revealed under the influence of VPA in C6 rat glioma models (Mean control 10,3 vs Mean VPA 14,8; Median control 11,0 vs Median VPA 13,0, days). It is noteworthy that the maximum life expectancy in the study group was 22 days, while in the control group it did not exceed 12 days

2. The Ki-67 proliferative index in the group where the VPA drug was used is significantly lower compared to the control group ( $2.17 \pm 0.38$ , Median=2.11 vs  $3.53 \pm 0.96$ , Median=3, 08)

3. Further studies of the oncostatic effect of VPA on glial tumors should be considered promising

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#### Funding

No funding was received for this research

#### Conflicts of interest/Competing interests

All authors certify that they have no affiliations with or involvement in any organization or entity with any financial interest (such as honoraria; educational grants; participation in speakers' bureaus; membership, employment, consultancies, stock ownership, or other equity interest; and expert testimony or patent-licensing arrangements), or non-financial interest (such as personal or professional relationships, affiliations, knowledge or beliefs) in the subject matter or materials discussed in this manuscript.

#### Availability of data and material

Not applicable

#### Code availability

Not applicable

#### Ethics approval

All applicable international, national, and/or institutional guidelines for the care and use of animals

were followed. The work was approved by the Bioethics Committee No. 43 dated 04.24.23

#### Consent to participate

Not applicable

#### Consent for publication

Not applicable

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